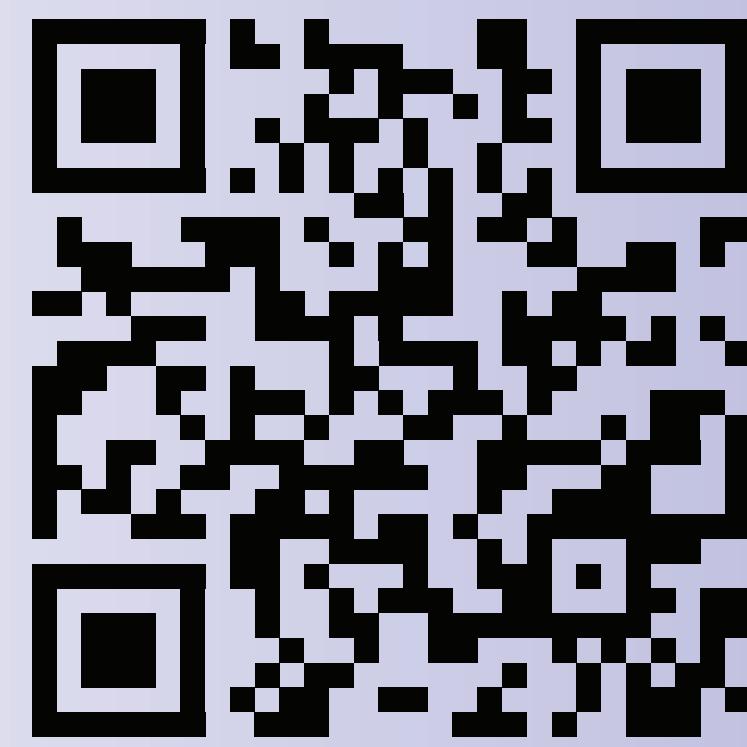


CHARACTERIZATION OF MEMBRANE INTERACTIONS WITH LACTOFERRICIN PEPTIDES BY ALL-ATOM AND COARSE-GRAINED MOLECULAR DYNAMICS SIMULATIONS, SOLID-STATE NMR, AND FLUORESCENCE SPECTROSCOPY



<http://tinyurl.com/6t92pb>

Tod D. Romo¹, Joshua N. Horn¹, Denise V. Greathouse², Alan Grossfield¹

¹University of Rochester Medical School, Rochester, NY, USA

²University of Arkansas, Fayetteville, AR, USA

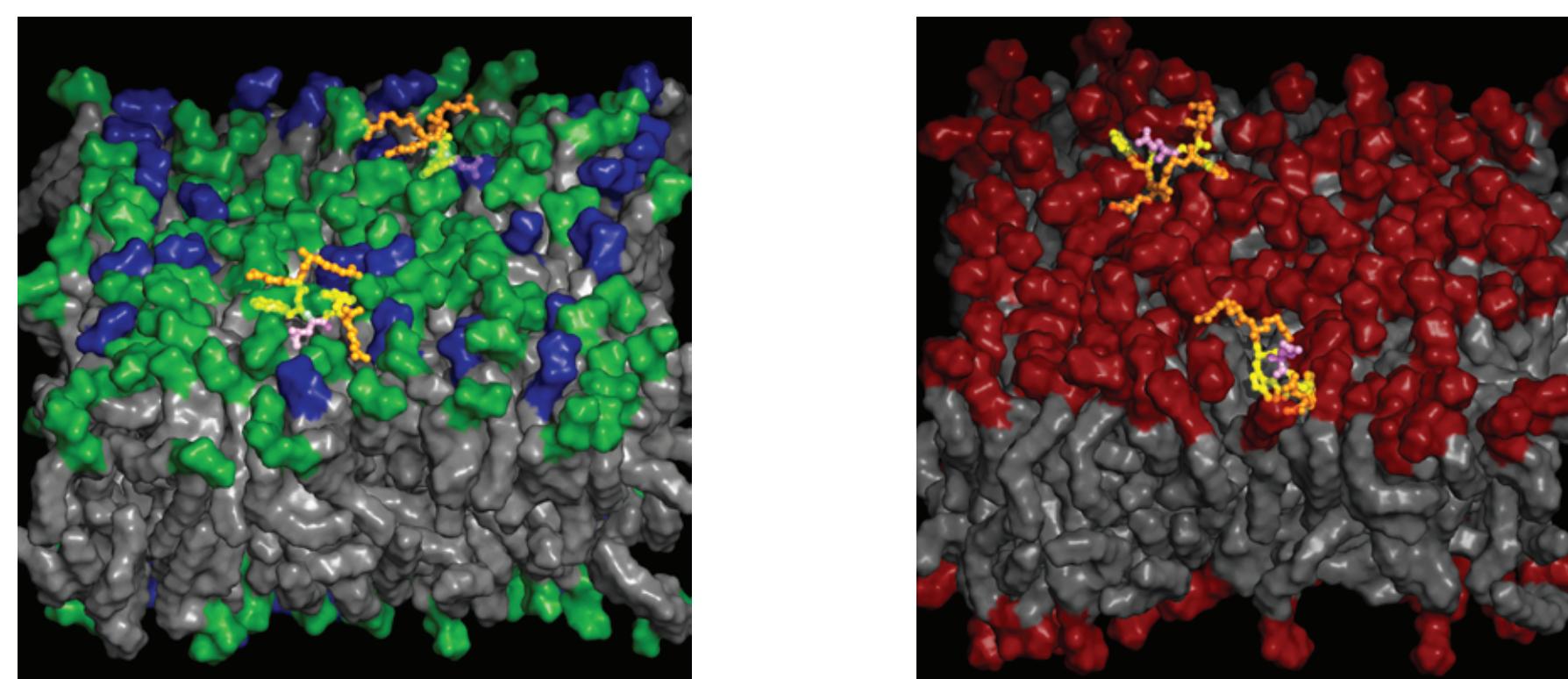


UNIVERSITY of
ROCHESTER

Abstract

LfB6 (RRWQWR-NH₂) is a small cationic antimicrobial peptide with broad spectrum effectiveness that is derived from bovine lactoferrin. The mechanism for interaction between the antimicrobial peptide and the bacterial cell membrane is hypothesized to depend on lipid composition. Bacterial membranes generally contain a significant fraction of negatively charged lipids in contrast with zwitterionic mammalian membranes. Previously, we characterized the interactions of an acylated LfB6 (C6-LfB6) with a model bacterial membrane (3:1 POPE:POPG) and a model mammalian membrane (POPC). Here, we investigate the interactions of the non-acylated LfB6 peptide with the same model membranes, using over 17 us of all-atom molecular dynamics as well as 53 μs of coarse-grained simulations, and we compare our results to solid-state ²H NMR and fluorescence spectroscopy. Molecular dynamics simulations reveal that the LfB6 peptide backbone does not penetrate as deeply in the model membranes as C6-LfB6 and that there is no preference in order of side-chain binding, unlike C6-LfB6. Further, molecular dynamics indicates the LfB6 tryptophans are more deeply buried in the membrane than C6-LfB6, yet fluorescence spectroscopy suggests they are more water-exposed. Coarse-grained molecular dynamics reveals that LfB6 comes off the membrane more easily than C6-LfB6, explaining the tryptophan membrane location and water exposure. The results also show subtle changes in the membranes' structure between the acylated and non-acylated peptides.

System Construction



All-Atom

- 2 peptides
- 100 lipids per leaflet
- POPE in green, POPG in blue
- Solvated to 50% w/w (7,900 waters)
- 50 mM salt (plus neutralizing)
- ~49,000 atoms
- CHARMM 27 forcefield
- Electrostatics using PME
- 10 Å vdW cutoff
- NP/T at 50°C

- 2 peptides
- 100 lipids per leaflet
- 2,000 waters
- 2,000 waters
- 50 mM salt (plus neutralizing)
- NPT at 50°C

MARTINI forcefield v2.1 with GROMACS 4.5.3 and 4.5.4

3:1 POPE:POPG

POPC

Simulations

All-Atom						Coarse Grained						
Membrane	Type	Tension (dyn/cm)	Length (nm)	Avg Length (nm)	Arcs / Lipid (Å)	Avg Arcs / Lipid (Å)	Membrane	Type	Length (nm)	Avg Length (nm)	Arcs / Lipid (Å)	Avg Arcs / Lipid (Å)
POPE:POPG	Neat	32.5	237	239	65.4	65.7	POPE:POPG	C6-LfB6	3100	64.9	63.6	63.6
			238		65.8			3417	64.9	63.6	63.6	
			528		65.5			3055	64.9	63.6	63.6	
			632		66.3			3002	64.9	63.6	63.6	
			630		65.6			3428	63.5	63.5	63.5	
			330		65.4			3406	63.5	63.5	63.5	
			345		65.1			3001	63.4	63.4	63.4	
			333		65.4			3002	63.4	63.4	63.4	
			231		65.3			3549	67.8	67.8	67.8	
			862		65.2			3600	67.8	67.8	67.8	
			4300		67.8			3102	67.8	67.8	67.8	
			1661		64.9			3400	67.7	67.7	67.7	
			1798		64.8			4400	67.6	67.6	67.6	
			348		70.4			3003	67.6	67.6	67.6	
			345		64.8			3012	67.7	67.7	67.7	
			353		70.0							
			585		70.5							
			672		71.1							
			664		71.1							
			632		71.1							
			1145		70.9							
			859		70.7							
			930		70.9							
			1190		70.8							

The first 100ns is considered equilibration and excluded from calculations

Methods

- 5 Å probe radius
- Count atoms within the sphere
- Fractional contribution by different components
- Peptide heavy atoms probed for entire peptide, all arginine atoms, all tryptophan atoms, and all C6 tail atoms (not shown)
- Time series averaged across all simulations

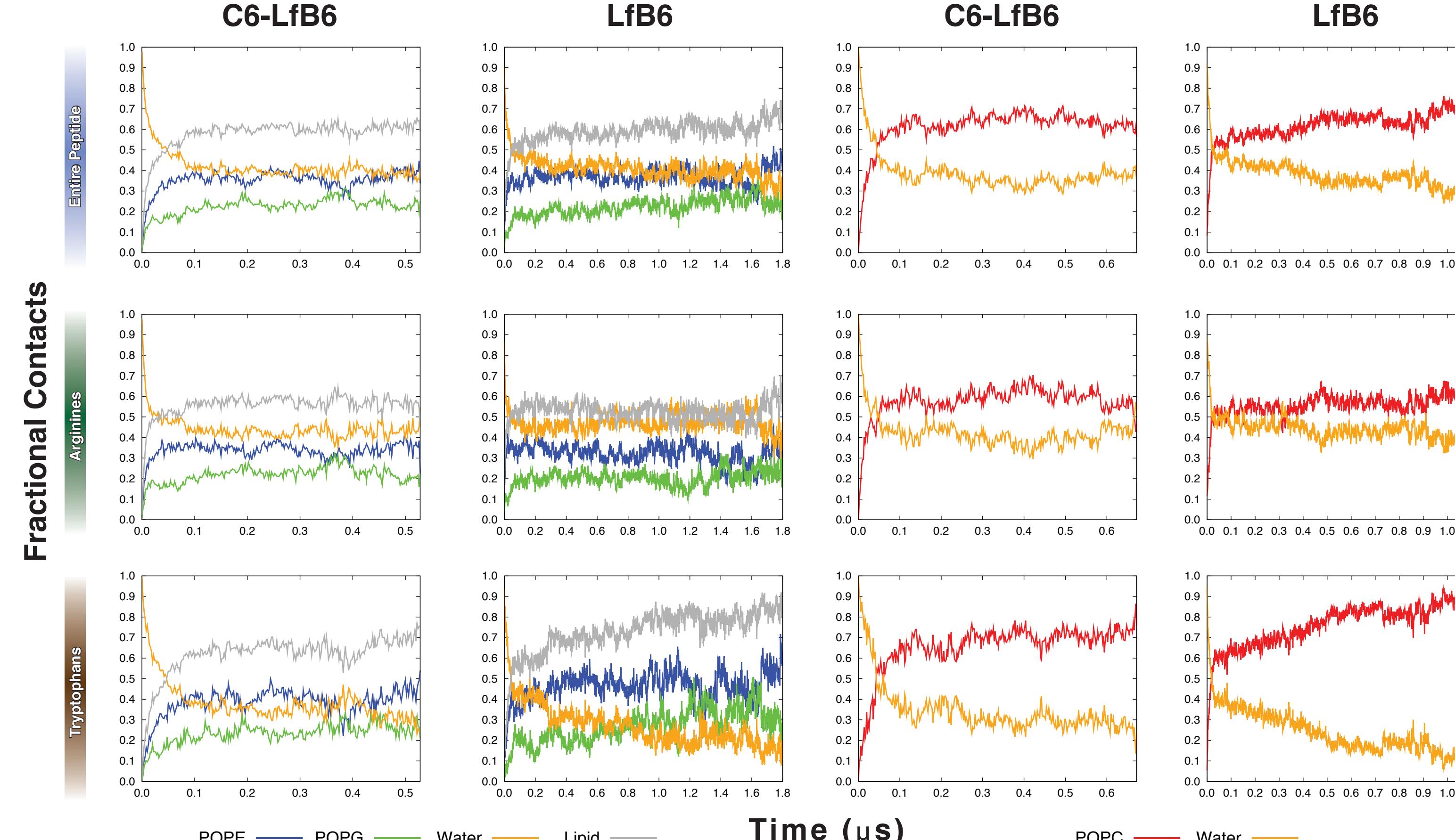
POPE:POPG

- Acylated:
 - Arg touches first, followed by Trp and then C6
 - Trp has slightly more lipid contacts than Arg
 - POPG contacts nearly equal POPE despite 3:1 ratio in membrane.
- Non-Acylated:
 - No order seen in contact
 - POPG contacts nearly equal POPE despite 3:1 ratio in membrane
 - Trp makes more lipid contacts than acylated

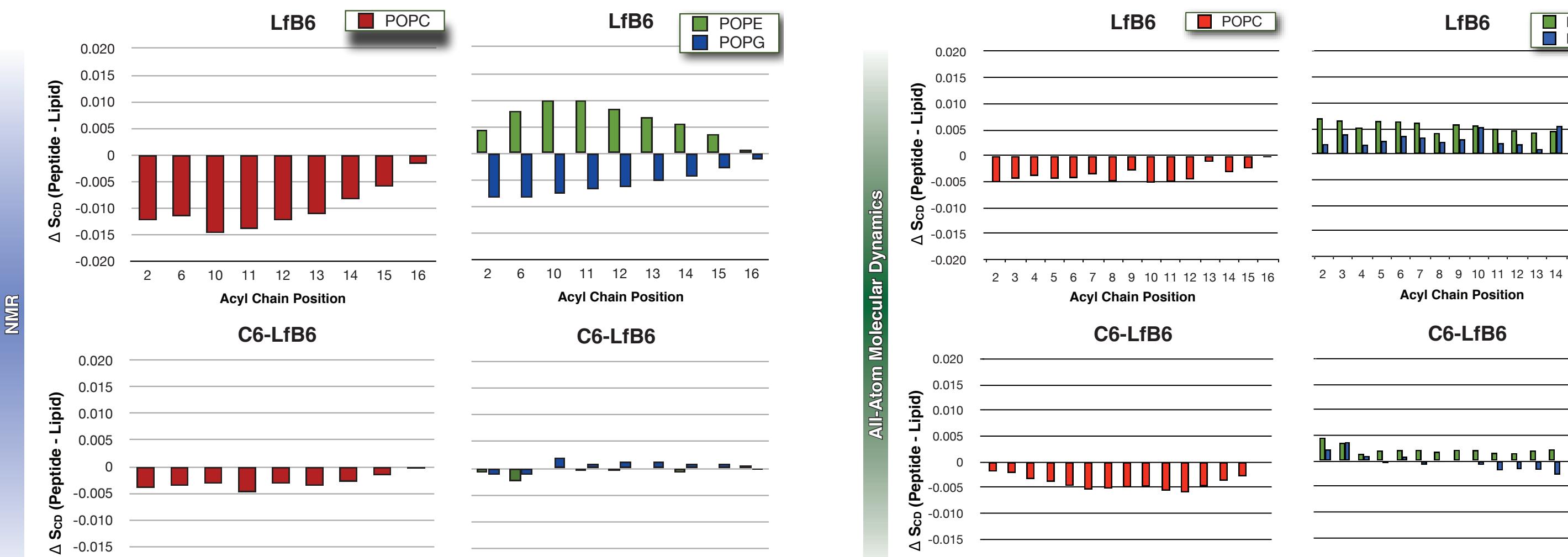
POPC

- Acylated:
 - C6 tails touch first (not shown), followed by Arg and Trp
 - Trp makes slightly more lipid contacts than acylated POPE:POPG
- Non-Acylated:
 - No preference in contact order
 - Trp makes more lipid contacts than acylated and non-acylated POPE:POPG

Lipopeptide Binding Mechanism



Effects on Membrane Structure: ²H Order Parameters



Distance-Based Order Parameters

Methods

- Simulation order parameters calculated using LOOS
- Acyl C-H bond orientation relative to membrane normal: $S_{CD} = -\frac{1}{2} \langle 3 \cos^2 \theta_{CD} - 1 \rangle$
- Experimentally measured by deuterium quadrupole splitting in solid state NMR

Discussion

- Subtle changes in membrane order for acylated peptide.
- Relative pattern of membrane order agrees between MD and NMR, despite differing absolute order parameters:
 - POPC < POPG < POPE

Discussion

- Peptide significantly decreases chain order at short range
- Slightly larger decrease in short range chain order for acylated peptide
- Larger decrease in POPC order, followed by POPG, and then POPE

Molecular Order Parameters

Methods

- Calculate principal components for lipids
- Use 2nd and 3rd principal components in lieu of C-H bond
- "Molecular Order Parameter" calculated as above
- Consider only lipids on the same leaflet as peptide
- "Molecular Order Parameter" binned based on lateral distance to nearest peptide

- Significant short-range effect on membrane order in all systems and peptides
- Acylated peptide decrease short-range order more than non-acylated
- POPC < POPG < POPE
- Good agreement between all-atom and coarse-grained molecular dynamics
- Origin of "hump" in coarse-grained data unclear

Peptide Water Exposure

Electron Density

Methods

- Electron density along the membrane normal is calculated using LOOS
- Membrane interface is defined as Z-axis location of peak lipid head group density
- Electron density for peptide backbone, tryptophans, and arginines plotted relative to membrane interface
- Density is normalized for visualization purposes

Discussion

- Backbone of acylated peptide resides deeper in both membranes than the non-acylated
- Backbone is buried more deeply in POPC than in POPE:POPG
- Trp is more deeply buried in POPC
- Arg remains near the membrane interface
- The acylated Arg is more buried than the non-acylated

Fluorescence Spectroscopy

Methods

- Trp emission fluorescence spectroscopy
- 1:50 peptide:lipid ratio

Discussion

- Non-acylated Trp in POPC is blue-shifted suggesting greater water exposure
- Lipid/water contacts and electron density plots indicate Trp is more deeply buried in the POPC membrane

Conclusions

- The acylated and non-acylated peptides both have subtle effects on the membrane
- The relative order is consistent between MD and NMR:
 - POPC < POPG < POPE
- Both peptides show significant membrane effects at short range
- Unlike the acylated peptide, the non-acylated peptide shows no preference in binding sequence
- The acylated peptide binds deeper in the membrane than the non-acylated one
- Tryptophans reside deeper in POPC membranes than POPE:POPG
 - Fluorescence suggests greater water exposure for Trp in POPC
- Coarse-grained MD shows that the non-acylated peptide comes off the POPC membrane, exposing the Trp to water
- Combining all-atom and coarse-grained MD reconciles the seemingly contradictory fluorescence and MD data
- Acylation increases the "stickiness" of the peptide

We thank the Center for Integrated Research Computing at the University of Rochester for providing the BlueGene/P. The NMR facility and mass spec facility are supported by NIH grant 1 P30 RR 031154. Funding for DVG was also provided by R01 6460 and the Arkansas Biosciences Institute.

LOOS (Lightweight Object Oriented Structure analysis library) is a project of the Grossfield Lab and is an open-source library using C++ and BOOST to provide an easy to use and easy to extend framework for rapidly developing analytical tools for molecular simulations. LOOS is available through SourceForge at: <http://loos.sourceforge.net>

